Sex-related response of mice to silver nanoparticles after sub-acute exposure

<u>Lucija Božičević (lbozicevic@imi.hr)</u>1, Rinea Barbir1, Walter Goessler2, Vedran Micek1, Željko Debeljak^{3,4}, Ivan Pavičić1, Marija Ćurlin⁵, Dunja Gorup⁵, Ivana Vinković Vrček1, Blanka Tariba Lovaković1

¹Institute for Medical Research and Occupational Health, Zagreb, Croatia; ²Institute of Chemistry, University of Graz, Graz, Austria; ³Department for Clinical Laboratory Diagnostics, Clinical Hospital Osijek, Osijek, Croatia; ⁴Faculty of Medicine, Josip Juraj Strossmayer University of Osijek, Osijek, Croatia; ⁵School of Medicine, University of Zagreb, Zagreb, Croatia

Results Introduction Kidney Liver Kidney Lung Liver Lung Brain Β Brain silver The of Α extensive use 0,80 nanoparticles (AgNPs) represents a intact vs. gonadectomised 0,80 KIDNEY: F(1,26)=5.9091, p<0.05 public concern in regards to safety 0,60 health risk (1). Their adverse and 0,60 effects include induction of oxidative (b) ssem (a) (0,40) stress and may be dependent on AgNPs biological fate and interaction

with biomolecules (2 - 8). This study aimed to investigate the differences sex-related the in biodistribution and oxidative stress response of adult mice after subacute exposure to low-dose of AgNPs. Intact and gonadectomised male and female mice treated were intraperitoneally with polyvinylpyrrolidone (PVP)-coated and transferrin (TRF)-coated AgNPs in a dose of 1 mg Ag/kg b.w. during 21 day.

Materials and methods

The total Ag concentrations in the freeze-dried tissue samples were measured with ICPMS after microwave-assisted acid digestion using an Agilent Technologies 7500cx ICPMS system (Agilent, Waldbronn, Germany).



Figure 1. Weights of brain, liver, kidney and lung in female (A) and male (B) mice (intact (I) and gonadectomised (G)) after 21 day exposure to PVP-AgNP and TRF-AgNP. Values represent mean and standard deviation of five rats per group (Main effects ANOVA (p<0.05)).



Figure 2. Total Ag levels in kidney, liver, lung and brain of female and male mice (intact (I) and gonadectomised (G)) after 21 day exposure PVP-AgNPs and TRF-AgNPs. Values represent mean (rhombus) and standard error (whisker) of five rats per group. Significant differences (p<0.05) between treated and control group are denoted with asterisk (*), and between TRF-coated versus PVP-coated AgNPs of the same sex and gonadectomy status with hashtag (#).

2.5E+06

5.0E+04

6,0E+05

Oxidative stress biomarkers in liver, kidneys, brain and lungs was measured by 2',7'-dichlorofluorescin diacetate (DCFH-DA), dihydroethidium (DHE) and monochlorobimane (MBCl) staining.

Statistical analysis of results was carried out using Dell Statistica 13.2 software (StatSoft, Tulsa, USA). Each group was composed of 5 animals. Normality of distribution was tested with the Kolmogorov-Smirnov test. The comparisons between the control and treated groups for the different measured variables were conducted using one-way ANOVA, followed by Tukey's HSD post hoc test when significant differences were found (p < 0.05).



Figure 3. Oxidative stress parameters measured in kidney (A), liver (B), lung (C) and brain (D) of female and male mice (intact (I) and gonadectomised (G)) after 21 day exposure to silver nanoparticles stabilized with PVP-coated (purple) or TRF-coated (dark blue) AgNPs. Values represent mean and standard error of five rats per group. Significant differences (p<0.05) between treated and control (green) samples are denoted with asterisk (*) and between TRF-coated versus PVP-coated AgNPs of the same sex and gonadectomy status with hashtag (#).

Conclusion

1,5E+05

Accumulation of Ag was significantly higher in the liver of females compared to males, as well as in the lungs of intact males compared to gonadectomised group. The effect of protein corona on AgNP accumulation was the most evident in brain of female mice. Sex-related differences were observed for ROS and GSH levels in almost all tissues. The highest difference was observed in lungs of female mice that responded more intensively to AgNPs exposure than males. AgNP distribution and toxicity are sex-related and dependent on the surface stabilization of AgNPs.

ACKNOWLEDGEMENT

This study received funding from the European Union's Horizon 2020 research and innovation program under grant agreement No. 814425 (RiskGONE – H2020–NMBP–13–2018 RIA).





References:

2,4E+05

1.5E+07

Li Y, Cummins E. Hazard characterization of silver nanoparticles for human exposure routes. J Environ Sci Heal - Part A Toxic/Hazardous Subst Environ Eng. 2020;55(6):704–25.
Korani M, Ghazizadeh E, Korani S, Hami Z, Mohammadi-Bardbori A. Effects of silver nanoparticles on human health. Eur J Nanomedicine. 2015;7(1):51–62.

3. Docea AO, Calina D, Buga AM, Zlatian O, Paoliello MMB, Mogosanu GD, et al. The effect of silver nanoparticles on antioxidant/pro-oxidant balance in a murine model. Int J Mol Sci. 2020;21(4).

4. Shrivastava R, Kushwaha P, Bhutia YC, Flora SJS. Oxidative stress following exposure to silver and gold nanoparticles in mice. Toxicol Ind Health. 2016;32(8):1391–404.

5. Barbir R, Goessler W, Ćurlin M, Micek V, Milić M, Vuković B, et al. Protein Corona Modulates Distribution and Toxicological Effects of Silver Nanoparticles In Vivo. Part Part Syst Charact. 2019 Aug;36(8):1900174.

6. Bertrand N, Grenier P, Mahmoudi M, Lima EM, Appel EA, Dormont F, et al. Mechanistic understanding of in vivo protein corona formation on polymeric nanoparticles and impact on pharmacokinetics. Nat Commun. 2017;8(1).

7. Garza-Ocanas L, Ferrer DA, Burt J, Diaz-Torres LA, Ramirez Cabrera M, Tamez Rodriguez V, et al. Biodistribution and lon-term fate of silver nanoparticles functionalized with bovine serum albumin in rats. Metallomics. 2010;2:204–10.

8. Barbir R, Goessler W, Ćurlin M, Micek V, Milić M, Vuković B, et al. Protein Corona Modulates Distribution and Toxicological Effects of Silver Nanoparticles In Vivo. Part Part Syst Charact. 2019;36(8).